

RESULTS OF RAPID ANTI MICROBIAL SENSITIVITY AND TOXICITY TEST OF THE EXTRACTS OF *SWERTIA CHIRATA*, *SYMPLOCOS RACEMOSA* AND *SOLANUM NIGRUM*

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ABSTRACT: An evaluation of anti microbial and cytotoxic effects of the plants extracts of *Solanum nigrum* (two varieties i.e. black and red fruits), *Swertia chirata*, *Symplocos racemosa* used as a medicinal agent in the cure of different ailment. The cytotoxic tests were carried ou~ on *Artemia Salina* (brine shrimp). The results showed that there was no positive lethality of any of the plant except the *Solanum nigrum* having black barrier/fruits (SNBS). LD50 i.e. 927.0293 j.lg/ml was found in upper toxic concentration, 258008.100 and lower toxic concentration is 231.0767.

The anti bacterial activity of crude extracts were studied *in vitro* on gram positive and gram negative bacteria *E. coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae* while antifungal activity was foundona range of concentration between 20-700mg/disc of dry extract and fungal organisms, *Candida albicans* and *Aspergillus niger*. The crude extracts except SNBS were devoid of any toxicity on *Artemia salina* with in the range of antimicrobial concentration, suggesting that the action is selective on microorganism.

INTRODUCTION

Swertia chirata (Gentianaceae) is an erect annual herb (Anonymous 1995). The drug consist chiefly of the stem, which is a dark purplish brown color, flower is paniced small green. It is widely used to treat fever, malaria and liver diseases (Najumdar and Guha, 1993). In addition it is reported to have anti-inflammatory activity. Xanthone deriva-tives are known to possess significant anti-inflammatory activities (Najumdar and Guha, 1993). Several varieties of xanthone show potent anti-platelet, anticancer, antifungal, antimalarial and significant CNS stimulant effects. It is also used as bitter antistomachic, febrifuge, antihelminthic, diuretic, antiepileptic and for certain type of mental disorders (Najumdar and Guha, 1993).

Symplocos racemosa (Symplocaceae/ Styraceae) is a small tree with dark green leathery leaves. In indigenous system of medicines *S. race1170sa* is used for management of menstrual disorders and to provide firmness to spongy and bleeding gums. Its decoction is also used for the treatment of bowel complaints and ulcer. The astringent bark of *S. racemosa* is recommended for the treatment of eye diseases and uterine disorders (Jain. 1985).

The leaves and fruits of *Solanum nigrum* L. (Solanaceae) i.e. black night shade locally called Mako, are used medicinally (Anonymous). Reported constituents of *S. nigrum* are solanine, saponin, oil, dihydroxy stearic, palmitic and stearic acid. The barriers are oleogenous, bitter, pungent, laxative, alterative, aphrodisiac, tonic and diuretic. They are 6mm in diameter, globoseusuaUy found in purplish black, green (unripe), red and sometime yellow. Different varieties of *S. nigrum* found in other part of the world included South Africa, Germany and America. It is used by Europeans as a remedy for convulsions. In Southern Rhodesia the plant is used as African remedy for malaria, black water fever, dysenteries and other diseases. In Mouriti!Js a poultice of the plant is applied for the relief of abdominal pain and inflammation of the urinary bladder. It has been also used in the treatment of headaches, ulcers and wounds (Akhtar and Muhammad, 1989; Jain and Borthakur,. 1986). The fresh. young leaves contain Img/100g of ascorbic acid. The frpit has been used as a tonic in heart diseases, fever, diarrhea (Anonymous 1956). eye disease and in liver and other complaints. Freshly prepared fluidextract from all portions of plant has been recommended dropsy. heart disease, piles. gonoIThea. inflammatory swellings (Watt and Breyer-Brandwijk, 1962) and chronic cirrhosis of the liver and spleen (Nadkami, 1976).

EXPERIMENT AL

Extraction of plant materials: *S. racemosa*, *S. chirata*

aemginosa (gram negative) and fungi *Candida albicans* and *Aspergillus niger* were obtained from Liaquat National Hospital Karachi. The bacterial cultures were maintained on a nutrient broth medium

Plants Name	<i>E. coli</i>	<i>K. pneumoniae</i>	<i>P. aeruginosa</i>	<i>S. aureus</i>
<i>S. chirata</i>	9mm	6mm	7mm	8mm
<i>S. racemosa</i>	7mm	6mm	6mm	8mm
SNBL	7mm	7mm	R	R
SNRL	R	9mm	8mm	8mm
SNRS	8mm	8mm	8mm	8mm
SNBS	7mm	9mm	8mm	6mm

Table 2 .
Antifungal Activity of Different Plant
Crude Extracts

Name of plant	Zone of inhibition against <i>Candida albicans</i>	Zone of inhibition against <i>Aspergillus niger</i>
<i>S. chirata</i>	R	9mm
<i>S. racemosa</i>	R	9mm
<i>S. nigrum</i> (SNBS)	8mm	R
<i>S. nigrum</i> (SNRS)	R	10mm
<i>S. nigrum</i> (SNBL)	8mm	9mm
<i>S. nigrum</i> (SNRL)	6mm	10mm

and two varieties of *Solanum nigrum* having red and black fruits were used to investigate the anti microbial and cytotoxic activities. Two kilograms of each plant material was crushed/cut into small pieces and soaked in ethanol (5 liters each) for 15 days. After this period material was filtered and the solvent was evaporated under reduced pressure on a rotary evaporator. A semisolid (gummy) extract was obtained. To this semisolid extract anti microbial and toxicity test were performed.

Antimicrobial activity of the crude extract: The pure cultures of bacteria *Escherichia coli* (gram negative), *Staphylococcus aureus* (gram positive) *Klebsiella pneumoniae* (gram negative), *Pseudomonas*

incubating at 37°C for 48 hours, while fungi used in antimicrobial activity were stored on Sabouraud Dextrose agar in the refrigerator at 4°C before use. Minimum inhibitory concentration (MIC) of extracts and pure compounds was performed by using standard protocols (Naqvi et al., 1992).

Cytotoxicity assay: Solutions of the test sample were prepared by initially dissolving the 10 mg in 1 ml of DMSO and from this solution transfer 500 µl, 50 µl, and 5 µl to vials (3 vials/conc.) corresponding to 1000, 100 and 10 µg/ml respectively. Dried it over night to conduct the brine shrimp lethality bioassay, maintained the brines shrimps eggs at 3TC in to the tank having continuous oxygen supply. Allow the eggs for maturation then place 30 larvae in to each vial containing 1000, 100 and 10 µg/ml of test samples. Observe the number of survived larvae in each vial (Meyer et al, 1982; Maclaughlin et al., 1990; Amason et al., 1989).

RESULTS AND DISCUSSION

The antibacterial activity of crude extract of *S. chirata*, *S. racemosa*, *S. nigrum* (SNBL SNRL SNRS. SNBS) were performed on gram +ve and gram -ve microorganism. The results of this activity were measured in term of zone of inhibition. *S. chirata* showed maximum inhibitory response against *E. coli*. moderate activity against other selected microorganism where as *S. racemosa* showed highest response against *S. aureus*. No inhibitory zone was recorded against *P. aeruginosa* by *S. chirata*, *S.*

racemosa and SNBL but SNRL, SNRS and SNBS have positive results. In the data maximum response of the crude extract of SNRL was observed against *K. pneumoniae*.

The antifungal activity of all six crude extract was observed against *Candida albicans* and *Aspergillus niger*. SNBS SNBL and SNRL showed positive results against *Candida albicans* where as all the crude extracts except SNBS have significant activity against *Aspergillus niger*.

These antimicrobial results are a positive sign for further evaluation of the chemical constituents and can help in the future investigation of new antimicrobial ~~~.

The toxicities of crude plant extracts to wards brine shrimps (*Artemia salina*) were performed. Brine shrimp larvae are commonly used for cytotoxicity assays (Pelka *et al.*, 2000). These larvae are sensitive to toxic substances (Sultana *et al.*, 1999). The ratio between dead larvae (high motility in comparison to a control with out any toxic substances is used to estimate the toxicity of the test solution. The results show that there is no positive lethality for any of the plant extract except the SNBS which shows the positive cytotoxic effect with LD₅₀ 927.0293 Ilg/ml (upper toxic conc. is 258008.100 and lower toxic concentration is 231.0767). This assay on medicinal plants is carried out to evaluate the anticarcinogenic or antitumor activity. This is the relatively new technique and has some advantage for toxicity testing restorative materials because it can be quickly carried out at low cost.

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